AllPure Plasmid MaxiPrep Kit



Cat. No. ABTGNA305-10

Storage: RNase A at -20°C for one year; others at room temperature (15-25°C) for one year

Description

AllPure Plasmid MaxiPrep Kit provides an efficient way to isolate high-quality plasmid DNA from \leq 500 ml LB or \leq 100 ml AllPure Plasmid Culture bacterial culture. DNA is bound to silica-based column. Unique formulated lysis buffer and neutralization buffer permit error-free visual identification of complete bacterial cell lysis and neutralization. In addition, this kit uses a simple method to remove endotoxin on column. The purified plasmid DNA is suitable for a variety of molecular biology applications, including restriction enzyme digestion, ligation, transformation, DNA sequencing, and transfection.

Kit Contents

Component	10 rxns
Resuspension Buffer (RB)	120 ml
Lysis Buffer (LB, Blue)	120 ml
Neutralization Buffer (NB, Yellow)	160 ml
ToxinOut Buffer (TB)	60 ml
Wash Buffer (WB)	25 ml
Elution Buffer (EB)	30 ml
RNase A (10 mg/ml) -20°C	1.2 ml
Maxi-Plasmid Spin Columns with Collection Tubes	10 each

Procedures

★ Prior to use, add RNaseA to RB, store at 4°C; add 100 ml of 96-100% ethanol to WB.

Bacterial cell culture (Lysogeny broth medium)	RB	LB	NB
≤100 ml	5 ml	5 ml	7 ml
100 ml-200 ml	10 ml	10 ml	14 ml
200 ml-300 ml	15 ml	15 ml	21 ml
300 ml-400 ml	20 ml	20 ml	28 ml
400 ml-500 ml	25 ml	25 ml	35 ml

1. Centrifuge overnight bacterial cell culture (refer to above table) at 12,000×g for 2 minutes and discard the supernatant.

2. According to the reference table, add 5~10 ml of colorless RB (containing RNase A). Mix thoroughly by vortexing.

- 3. According to the reference table, add 5~10 ml of blue LB, mix by gently inverting the tube 5-6 times to completely lyse cells. Incubate at room temperature for 5 minutes (lysate should change color to blue).
- 4. According to the reference table, add 7~14 ml of yellow NB, mix by gently inverting the tube 5-6 times. Incubate at room temperature for 2 minutes (lysate should change color to yellow).
- 5. Centrifuge at 12,000×g for 15 minutes, gently transfer the supernatant to a spin column (if more than 50 ml, repeat the transfer after step 6).
- 6. Centrifuge at 8,000×g for 2 minutes and discard the flow-through.

7. Add 5 ml of TB, incubate at room temperature for 10 minutes. Centrifuge at 8,000×g for 2 minutes and discard the flow-through.

8. According to the reference table, add 3~5 ml of WB, centrifuge at 8,000×g for 2 minutes and discard the flow-through.

9. Repeat step 8 once.

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- 10. Centrifuge at 8,000×g for 5 minutes to completely remove the remaining WB.
- 11. Incubate at room temperature for 10 minutes to evaporate ethanol.
- 12. Place the spin column in a clean 50 ml centrifuge tube, add 1-2 ml of of EB or sterile, distilled water (pH >7.0) directly to the center of the column matrix (for higher yield, use prewarmed (60-70°C) EB or distilled water).
- 13. Centrifuge the column at 8,000×g for 2 minutes to elute DNA.
- 14. Isolated plasmid DNA can be stored at -20°C Optional (to futher concentrate DNA)

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- 1. Transfer eluent to a microcentrifuge tube, add 1/10 volume of NaAC (3 M, pH 5.2) and 7/10 volume of isopropanol (at room temperature). Mix thoroughly and incubate at room temperature for 5 minutes.
- 2. Centrifuge at 12,000×g for 10 minutes and discard the supernatant.
- 3. Add 1 ml of 70% ethanol (room temperature), centrifuge at 12,000×g for 10 minutes and discard the supernatant.
- 4. Air-dry the pellet for 5-10 minutes. Add appropriate volume of EB to dissolve the pellet

Notes

- All centrifugation steps are carried out at room temperature.
- AllPure Plasmid Culture is recommended for growing low-copy number plasmid.
- After adding LB or NB, don't mix by vortexing. Vigorous mix may result in genome contamination.
- Add the whole volume of RNase A (supplied with this kit) into RB solution, mix thoroughly and store at 4°C.
- Prior to use, check whether the LB is cloudy or not, if it is cloudy, heat it in 37°C water bath to completely dissolve it. Close the cap immediately after each use to avoid pH change.
- Make sure to use the right cell culture volume. Too much cell culture can result in incomplete lysis, which will affect plasmid DNA yield and purity.

Prior to use, add RNaseA to RB Centrifuge at 8,000×g for 2 minutes Bacteria and discard the flow-through. Cell Harvesting add RB RB Mix by Bacterial cell culture votexing (Lysogeny broth medium) WB Add 3~5 ml of WB LB add LB Repeat step 8 once Invering the tube 4~6 times to completely lyse cells (lysate become to blue color) Centrifuge at 8,000×g for 2 minutes and discard the flow-through. NB add NB Invering the tube 5~6 times Incubate 2 mins at RT (lysate become to yellow color) Incubate at RT for 10 minutes Centrifuge at 12,000×g for 15 minutes Place the spin column in a clean 50 ml centrifuge tube Spin Column Gently transfer the supernatant to a spin column add 1-2 ml of EB or EB sterile, distilled water (pH >7.0) Centrifuge at 8,000×g for 2 minutes Centrifuge the column at $8,000 \times g$ for 2 minutes to elute DNA. and discard the flow-through Add 5 ml of TB ΤB Incubate at RT for 10 minutes Isolated plasmid DNA can be stored at -20°C Optional (to futher concentrate DNA) FOR RESEARCH USE ONLY 3

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